







Exploring the Therapeutic Promise of Naphthoquinones: A Spotlight on Lapachol

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Received: 1.05.2024; Accepted: 6.10.2024; Published: 30.03.2026

Abstract: Medicinal plants possess various biological activities and significantly contribute towards the development of various clinically used medications. In recent years, the research interest in natural drug designs has increased exponentially. Lapachol is a useful phytoconstituent with a quinone ring, reported for its broad-spectrum pharmacological activities, including antimicrobial, anticancer, anti-leishmanial, anti-trypanosomal, molluscicidal, wound-healing, and other activities. The therapeutic values of Lapachol from various plant families have been explored in existing scientific reports, as well as in traditional uses and other potential therapeutic effects. This review highlights updates on the pharmacological activities and other traditional uses of Lapachol.

Keywords: antileishmanial; lapachol; molluscicidal; side effects; traditional.

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1. Introduction

The quinone moiety has attracted interest in recent decades due to its diverse pharmacological actions. In 1946, Wendel WB reported that some of the 2-hydroxy-3-alkylnaphthoquinones could inhibit the growth of *Plasmodium*. Quinone compounds generally have a p-bisdienonic ring system in their molecular structure and possess the capacity for reversible chemical reactivity toward oxidation-reduction [1]. Naturally occurring quinones and naphthoquinones are concerned with photosynthesis and are widely distributed in the lower as well as higher species of plants, animals, fungi, etc [2,3].

Lapachol was identified as the most abundant quinone in the family Bignoniaceae and, for the first time, isolated from *T. avellanae* in 1882 [4]. In Brazil, Lapachol is known as Pau D'Arc, Ipe-Black, Ipe-purple, and Yellow-Ipe [5]. The blood-brain barrier crossing of lapachol has been demonstrated in various in vivo studies and has been found effective in the treatment of malignant glioma [6]. Lapachol has been traditionally used in Brazil to treat malaria and fever. The antimalarial drug atovaquone, synthetically known as 2-alkyl-3-

hydroxynaphthoquinone, is also an analog of lapachol [7,8]. Other pharmacological activities of Lapachol are listed in Table 1 and diagrammatically represented in Figure 1.

Table 1. Pharmacological activities of Lapachol.

Sr. No.	Disease	Pharmacological activity	References
1	Cancer	Antitumor/anticancer/ antimetastaic activity	[9-15]
2	Chagas' disease	<i>In vitro</i> inhibition of amastigotes of <i>L. viannia braziliensis</i> and <i>Trypanosoma cruzi</i>	[16,17]
3	Disease in fish and mammals	Molluscicidal	[18]
4	Fungal infection	Antifungal	[19]
5	Hepatic cirrhosis	Hepatoprotective	[20]
6	Inflammation	Anti-inflammatory	[21]
7	Leishmania	Antileishmanial	[16-22]
8	Malaria	Antimalarial (<i>Ae. aegypti</i>)	[23,24]
9	Microbial infection	Antimicrobial	[25,26]
10	Schistosomiasis	Cercaricidal and schistosomicidal	[27]
11	Ulcer	Antiulcer	[28]
12	Viral disease	Antiviral	[29]
13	Blood clotting	Vitamin K-antagonist antigen	[30]

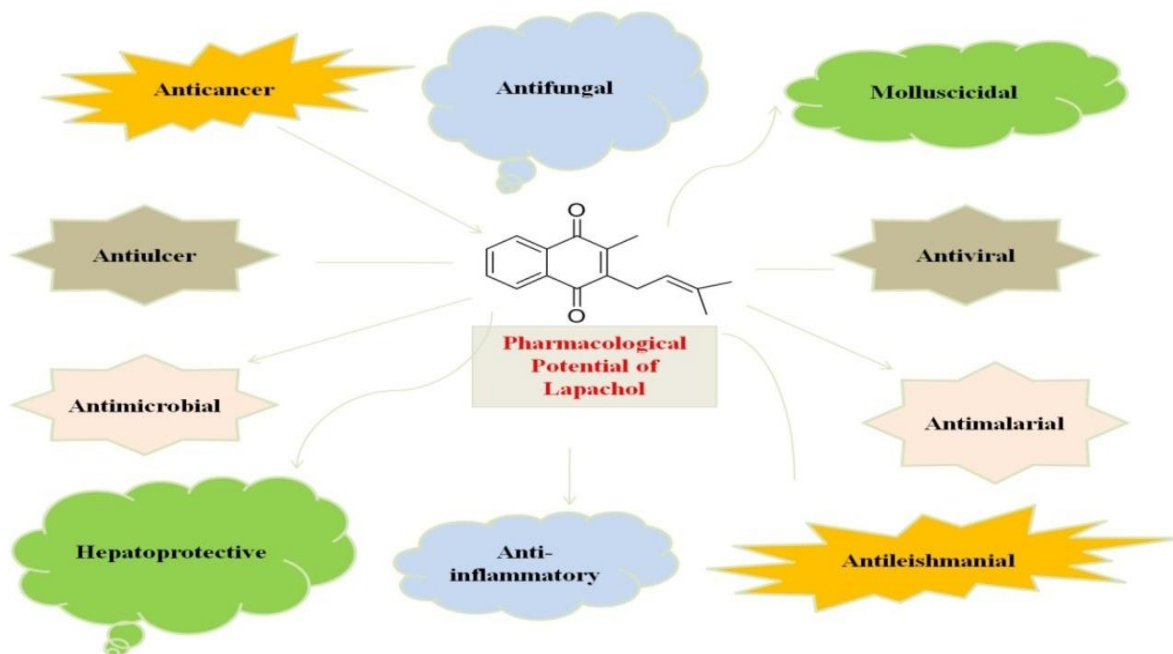


Figure 1. Pharmacological potential of Lapachol.

2. Biological Sources of Lapachol

Lapachol is generally found in bacteria, fungi, and higher plants. Lapachol is mainly distributed in *Bignoniaceae*, *Verbenaceae*, *Proteaceae*, *Leguminosae*, *Sapotaceae*, *Scrophulariaceae*, and *Malvaceae* [31]. It is mainly obtained from the plant extract of *T. avellanadae*, Taigu or lapachol wood, Teak wood, and root [32,33]. Some other biological sources of Lapachol are listed in Table 2.

Table 2. Various biological sources of Lapachol.

Sr. No.	Source	Family	Reference
1	<i>Tabebuia avellanadae</i>	Bignoniaceae	[32,33]
2	<i>Penicillium notatum</i>	Trichocomaceae	[34]
3	<i>Tectona grandis</i>	Lamiaceae	[32,35,36]
4	<i>Tabernaemontana undulata</i>	Bignoniaceae	[37,38]
5	<i>Tecoma stans</i>	Bignoniaceae	[39,40]
6	<i>Avicennia marina</i>	Acanthaceae	[41]
7	<i>Radermachera xylocarpa</i>	Bignoniaceae	[42]

Sr. No.	Source	Family	Reference
8	<i>Tabebuia rosea</i>	Bignoniaceae	[43]
9	<i>Tabebuia serratifolia</i>	Bignoniaceae	[43]
10	<i>Plumbago zeylanica</i>	Plumbaginaceae	[44]
11	<i>Kigelia pinnata</i>	Bignoniaceae	[45]
12	<i>Aspergillus niger</i>	Trichocomaceae	[46]
13	<i>Alternaria alternate</i>	Pleosporaceae	[46]
14	<i>Heterophragma adenophyllum</i>	Bignoniaceae	[47]
15	<i>Acacia nilotica</i>	Leguminosae	[48]

3. Chemical Properties

Chemically, Lapachol is 2-hydroxy-3-(3-methylbut-2-enyl)naphthalene-1,4-dione (Figure 2), a yellow color natural pigment, having chemical formula C₁₅H₁₄O₃, weakly acidic, low solubility in water and highly soluble in basic solutions [49-51].

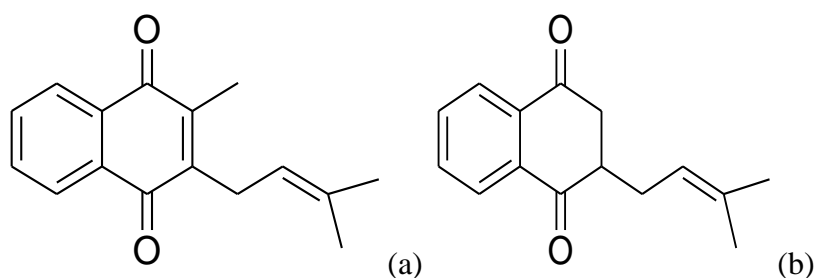


Figure 2. Chemical structures of lapachol (a) and dehydroxylapachol (b).

Various techniques are available for detecting Lapachol in aqueous solutions, blood, urine, and other biological fluids. In 1994, Alcanfor *et al.* determined lapachol in aqueous dispersion using a fluorescent probe 3MPA-CdTe QDs. Lapachol was reduced to the fluorescent product by sodium hydrosulfite, and this method could be used to verify lapachol in serum and pharmaceutical formulations with a limit of detection (LOD) in the g L⁻¹ range [52,53]. In 1995, Steinert *et al.* used a combination of HPLC and UV spectrometry for the detection of naphthoquinones in plant extracts, but were unsuccessful in identifying lapachol [54]. In 2004, Fonseca *et al.* recently validated lapachol in lapachone by using absorption detection and isocratic elution at 278 nm with an LOD of 0.3 g L⁻¹ in HPLC [55]. In 2013, Aucélio RQ *et al.* utilized the Stern-Volmer Luminescent semiconductor quantum dots (QDs) dispersions model to identify a relationship between the concentration of lapachol and fluorescence. The 3MPA-CdTe QD probe was also shown to be effective for the identification of lapachol in urine samples [56].

4. Potential Pharmacological Activities of Lapachol

Lapachol quinone is present in various plants and fungal species and has been known since ancient times for its antiprotozoal activity against malaria. However, Lapachol shows strong inhibitory activity against leukemic cells, protozoans, and fungal cells.

4.1. Antileishmanial activity of Lapachol.

In 2012, Ali A. determined the anti-leishmanial activity of dichloromethane and *n*-hexane fractions of *T. avellaneda*, and the earlier fraction has higher activity against extra and intracellular *L. major* parasites as compared to the latter one. This fraction of *T. avellaneda* was found to contain lapachol and a mixture of furanonaphthoquinones (5-hydroxy and 8-hydroxy-2-(1'-hydroxyethyl) naphtho[2,3-b]furan-4,9-dione (Figure 3). So, Lapachol might

be responsible for the antileishmanial activity of the plant extract, and the mechanism may involve ROS [57,58].

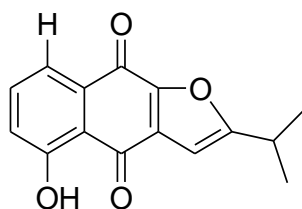


Figure 3. Structure of furanonaphthoquinone.

4.2. Anti-trypanosomal activity of Lapachol.

In 2000, Pinto *et al.* evaluated the biological activities of twenty-two lapachol heterocyclic derivatives against *T. cruzi*. Out of these, the oxazolic and imidazolic derivatives (Figure 4) were found to show 1.5 to 34.8 times superior activity than standard, i.e., crystal violet. These results indicate that these derivatives could be used as lead molecules for the treatment of Chagas disease [59].

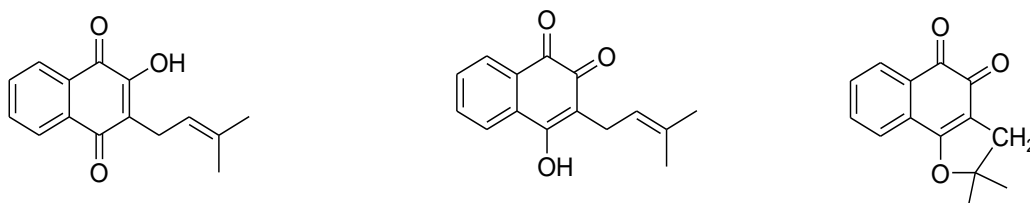


Figure 4. Oxazolic and imidazolic derivatives of lapachol.

In 2002, lapachol derivatives were tested against blood infected with trypomastigote (*T. cruzi*); triacetoxo derivatives were found to have better trypanocidal activity. As lapachol can be readily extracted and its derivatives are readily synthesized, large-scale field tests can be conducted for its applications in large-scale molluscicidal programs [60].

4.3. Antimicrobial activity of Lapachol.

In 2006, Park *et al.* reported the growth-inhibitory activity of *Tabebuia impetiginosa* (bark) phytoconstituents against *Helicobacter pylori* (ATCC 43504) using the MIC and paper disc diffusion methods. Standard antibiotics amoxicillin, metronidazole, and tetracycline were used for reference standards. The active components characterized in *Tabebuia impetiginosa* bark extract include anthraquinone-2-carboxylic acid, 2 (hydroxymethyl) anthraquinone, and 2-hydroxy-3-(3 methyl-2-butenyl)-1,4-naphthoquinone (Lapachol). Here, the activity of 1,4-naphthoquinone derivatives (lapachol) against *Helicobacter pylori* was greater than that of metronidazole [61].



Figure 5. Thiosemicarbazone and semicarbazone derivatives of Lapachol.

Moreover, in 2013, Souza *et al.* reported the antimicrobial activity of semicarbazone and thiosemicarbazone derivatives (Figure 5) of lapachol against the bacterial sps. *Staphylococcus aureus*, *Enterococcus faecalis*, and yeast *Cryptococcus gattii*. The thiosemicarbazone derivative of Lapachol was found to be active against eleven clinical isolates of *Paracoccidioides brasiliensis*. The MIC values of thiosemicarbazone derivatives of Lapachol against *S. aureus*, *P. brasiliensis*, *C. gattii*, and *E. faecalis* indicate that these compounds have potential in the development of novel drugs for the treatment of microbial infections [62].

4.4. Antifungal activity of Lapachol.

The antifungal potential of Lapachol is demonstrated in Table 3.

Table 3. Antifungal potential of Lapachol.

S. N.	Source of Lapachol	Fungal strains used for testing	Standard	Mechanism involved	References
1	Species of <i>Tabebuia</i>	<i>C. albicans</i> , <i>C. tropicalis</i> and <i>Cryptococcus neoformans</i>	Ketoconazole	Uncoupling oxidative phosphorylation and electron transport inhibition	[63,64]
2	Endophyte extracts of <i>A. niger</i> and <i>A. alternata</i>	Different pathogenic bacteria and fungi	Bavistin	-----	[65]
3	1,4-naphthoquinone derivatives of Lapachol	<i>S. aureus</i>	Vancomycin	-----	[19]

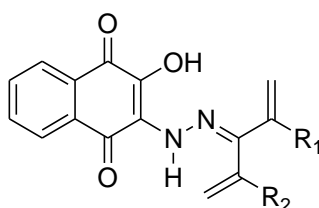


Figure 6. Chemical structure of hydrazone 1,4-naphthoquinone derivatives of Lapachol.

4.5. Molluscicidal activity of Lapachol.

Some studies reported that partially hydrogenated Lapachol derivatives have significantly higher molluscicidal activity than Lapachol itself [18,65]. Sant'Ana A.E.G. *et al.* in 2002 found out the action of Lapachol, its potassium salt, and acetyl derivative against the snail *Biomphalaria glabrata* and its egg masses. The LD₅₀ values of Lapachol and its salts indicate a strong molluscicidal activity [60].

4.6. Antiviral activity of Lapachol.

Lapachol's antiviral activities are illustrated in Table 4.

Table 4. Antiviral activity of Lapachol.

S.N.	Lapachol source	Enzyme or viral species against which active	Mechanism	References
1	Lapachol itself	β -Lapachone inhibits eukaryotic DNA-dependent DNA polymerase- α	Redox potential	[66]
2	Lapachol itself	<i>Herpes virus</i> hominis types I and II, poliovirus, influenza virus, and vesicular stomatitis virus	-----	[67]
3	1,2-naphthoquinone derivatives	Should be replaced with Lapachol, a 1,4-naphthoquinone compound that has been	Interferon production or	[68]

S.N.	Lapachol source	Enzyme or viral species against which active	Mechanism	References
		demonstrated to inhibit Nsp9 from SARS-CoV-2, which could be an important target in prospecting for ligands with antiviral potential	enzyme inhibition	
4	Lapachol	inhibition of vitamin K epoxide reductase and vitamin K quinone reductase	-----	[69]
5	Aq. Extracts of <i>T. avellanedae</i> inner bark	Epstein-Barr virus (EBV)	-----	[70]

4.7. Anticancer activity of Lapachol.

Interaction of Lapachol with nucleic acids of the DNA helix leads to inhibition of DNA replication and RNA synthesis, and its interaction with topoisomerase enzymes is known for its anticancer mechanism [31, 71-76]. Molecular modeling studies showed that Lapachol and 2-(1-hydroxyethyl)-naphtho[2,3-b]furan-4,9-dione were active against human estrogen receptor alpha, and this finding could be used in the treatment of breast cancer [77]. Different anticancer studies of Lapachol are shown in Figure 7.

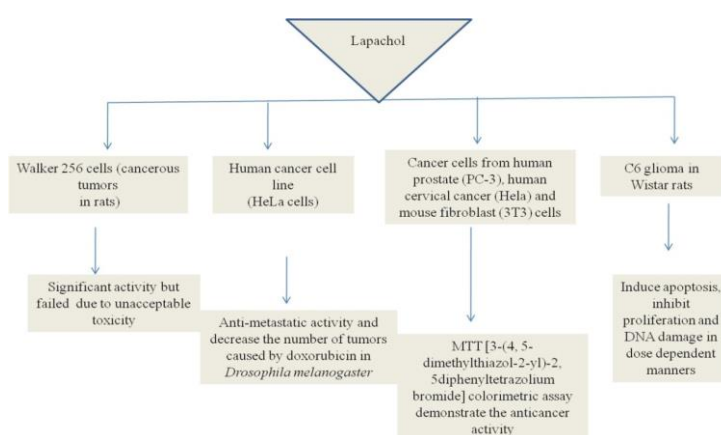


Figure 7. Different anticancer studies of lapachol.

A novel ruthenium(II) complex, along with lapachol, could also induce G2/M phase arrest by downregulating the Aurora-B kinase and downregulating the ROS-mediated apoptosis process in human prostate adenocarcinoma cells [78]. Lapachol's anticancer activity is shown in Table 5.

Table 5. Anticancer activities of lapachol.

S.N.	Source	Type of tumor or cells	Activity	References
1	Lapachol	Walker 256 cells (cancerous tumors in rats)	Significant activity and clinical trials were run by NCI in 1970, but they had to terminate them due to unacceptable levels of toxicity	[11,12]
2	Lapachol	Human cancer cell line (HeLa cells)	Anti-metastatic activity and decrease the number of tumors caused by doxorubicin in <i>Drosophila melanogaster</i>	[74]
3	Lapachol (methanolic extract of <i>Kigelia pinnata</i> root and its derivative)	Cancer cells from human prostate (PC-3), human cervical cancer (HeLa), and mouse fibroblast (3T3) cells	MTT [3-(4, 5-dimethylthiazol-2-yl)-2, 5-diphenyltetrazolium bromide] colorimetric assay demonstrate the anticancer activity	[79-81]
4	Lapachol	C6 glioma in Wistar rats	Induce apoptosis, inhibit proliferation, and cause DNA damage in a dose-dependent manner	[82]

4.8. Anti-inflammatory.

Lapachol shows anti-edematogenic activity by carrageenan pedal injection in rats. Initially, carrageenan shows a good anti-inflammatory activity in rats [21, 83-85].

4.9. Wound healing.

Wounds have been treated with natural substances for thousands of years [86, 87]. After isolating lapachol from *Fernandoa adenophylla*, Rauf *et al.* reported the drug's encouraging anti-inflammatory properties [88]. In 2023, Thiago Santos also reported the wound healing properties of Lapachol [89]. The importance of sirtuins in diabetic wound healing was illustrated by Bibi *et al.* in 2021, who identified a novel possible sirtuin activator, lapachol [90]. Lapachol may have inhibited extracellular matrix hyperplasia during wound healing and potentially reduced the development of hypertrophic scar, according to research by Matsui *et al.* [91].

In 2010, Sharma *et al.* reported the wound-healing activity of the aqueous extract of *K. pinnata* by the excision wound model. Aqueous extract of the bark of *K. pinnata* was found to contain a high amount of free ferulic acid, Lapachol, stigmasterol, and β -sitosterol, which implies that β -sitosterol may be responsible for the epithelization activity due to the free radical scavenging action [92]

4.10. Toxicity.

Orally administered lapachol also produced toxic effects in animals, with oral LD50 values of 0.621 g/kg and 2.4 g/kg in BALB/c mice and albino rats, respectively. Symbols of toxicity in dogs and monkeys include moderate to severe anemia, normoblastosis, reticulocytosis, elevated serum alkaline phosphatase activity, leukocytosis, and prothrombin times [93]. Some research results also indicate that Lapachol is not toxic to mothers, although it leads to fetal growth retardation and does not interfere with embryonic development, as well as the pre-implantation period. However, when administered during the organogenic period at the same dose level, it induced high fetal death incidences [94].

4.11. Snake bite.

Lapachol and synthetic derivatives were evaluated *in vitro* and *in vivo* against snake venoms of *Bothrops*. Natural quinone and other related synthetic quinones showed antivenom activity and reduced the inflammatory effects produced by venom. This pharmacological activity could be used in snakebite venom research and improve the management of cases of venomous snakebites [95-97].

5. Conclusion

As the currently available drugs are associated with various adverse effects, the current global attention to traditional medicine has led to rapid growth in the remedies used by various ethnic groups worldwide. The present article highlights the pharmacological potential of Lapachol, including antimicrobial, anticancer, antileishmanial, antitrypanosomal, molluscicidal, and wound-healing activities. So, Lapachol has the potential for further evaluation in animals and humans.

Author Contributions

Writing – review & editing, A. L., N. J., T. H., S. K., V. K., M. K. G., and N. A. All authors have read and agreed to the published version of the manuscript

Institutional Review Board Statement

Not applicable.

Informed Consent Statement

Not applicable.

Data Availability Statement

Not applicable.

Funding

This research received no external funding.

Acknowledgments

The authors would like to thank MM College of Pharmacy, Maharishi Markandeshwar (Deemed to be) University, Mullana, Ambala, for their kind support during the literature survey and all other data collection.

Conflicts of Interest

The authors declare no conflict of interest.

References

1. Wendel, W.B. The influence of naphthoquinones upon the respiratory and carbohydrate metabolism of malarial parasites. *Fed. Proc.* **1946**, *5*, 406-407.
2. Santra, I.; Mukherjee, S.; Haque, S.M.; Ghosh, B. Occurrence and Diversity of Major Naphthoquinones in Higher Plants: Their Distribution and Conservation Strategies. In: Jha, S., Halder, M. (eds) Medicinal Plants: Biodiversity, Biotechnology and Conservation. Sustainable Development and Biodiversity, Springer, Singapore **2023**, *33*, https://doi.org/10.1007/978-981-19-9936-9_15.
3. Spyroudis, S. Hydroxyquinones: Synthesis and Reactivity. *Molecules* **2000**, *5*, 1291-1330, <https://doi.org/10.3390/51201291>.
4. Hussain, H.; Krohn, K.; Ahmad, V.U.; Miana, G.A.; Green, I.R. Lapachol: An overview. *ARKIVOC* **2007**, *2*, 145-171, <http://dx.doi.org/10.3998/ark.5550190.0008.204>.
5. Almeida, E.R. Plantas Mediciniais Brasileiras [Medicinal Plants Brazilian popular and scientific knowledge], 1st Edition; Hemus, L.T.D.A., Ed.; São Paulo SP Brasil: **1993**.
6. de Souza-Fagundes, E.M.; Delp, J.; Prazeres, P.D.M.; Marques, L.B.; Carmo, A.M.L.; Stroppa, P.H.F.; Glanzmann, N.; Kisitu, J.; Szamosvári, D.; Böttcher, T.; Leist, M.; da Silva, A.D. Correlation of structural features of novel 1,2,3-triazoles with their neurotoxic and tumoricidal properties. *Chem Biol Interact* **2018**, *291*, 253-263. <https://doi.org/10.1016/j.cbi.2018.06.029>.
7. Oliveira, A.B.; Dolabela, M.F.; Braga, F.C.; Jácome, R.L.R.P.; Varotti, F.P.; Póvoa, M.M. Plant-derived antimalarial agents: new leads and efficient phythomedicines. Part I. Alkaloids. *An. Acad. Bras. Ciênc.* **2009**, *81*, 715-740, <https://doi.org/10.1590/s0001-37652009000400011>.
8. Looareesuwan, S.; Chulay, J.D.; Canfield, C.J.; Hutchinson, D.B. Malarone (atovaquone and proguanil hydrochloride): a review of its clinical development for treatment of malaria. Malarone Clinical Trials Study Group. *Am. J. Trop. Med. Hyg.* **1999**, *60*, 533-541, <https://doi.org/10.4269/ajtmh.1999.60.533>.
9. Kandioller, W.; Balsano, E.; Meier, S.M.; Jungwirth, U.; Göschl, S.; Roller, A.; Jakupec, M.A.; Berger, W.; Keppler, B.K.; Hartinger, C.G. Organometallic anticancer complexes of lapachol: metal centre-dependent formation of reactive oxygen species and correlation with cytotoxicity. *Chem. Commun.* **2013**, *49*, 3348-3350, <https://doi.org/10.1039/C3CC40432C>.

10. Subramanian, S.; Ferreira, M.M.C.; Trsic, M. A Structure Reactivity Relationship Study of Lapachol and Some Derivatives of 1,4-Naphthoquinones Against Carcino Sarcoma Walker 256. *Struc. Chem.* **1998**, *9*, 47-57.
11. Suffness, M.; Douros, J. Miscellaneous natural products with antitumor activity. In *Anticancer Agents Based on Natural Product Models*. Cassady, J.M., Douros, J.D., Eds.; Academic Press: New York, **1980**; pp. 474.
12. Rao, K.V.; McBride, T.J.; Oleson, J.J. Recognition and evaluation of Lapachol as an antitumor agent. *Cancer Res* **1968**, *28*, 1952-1954.
13. Kumar, A.; Sharma, S.; Kaushik, C.; Dhobal, M.P.; Joshi, B.C. Antitumor activity of Lapachol isolated from *Zizyphus nummularia* as adjuvant for radiation therapy. Tehran: Traditional medicines & Materia Medica Research Center, Shahid Beheshti University of Medical Sciences **2002**, *1*, 129-136.
14. Silva, M.N.; Vitor, F.; Souza, M.C.B.V. An overview of the chemistry and pharmacology of naphthoquinones with emphasis on beta-lapachone and derivatives. *Química Nova*, Conference: 26th Annual Meeting of the Sociedade-Brasileira-de-Quimica **2003**, *26*, 407-416.
15. Balassiano, I.T.; De Paulo, S.A.; Henriques Silva, N.; Cabral, M.C.; da Gloria da Costa Carvalho, M. Demonstration of the lapachol as a potential drug for reducing cancer metastasis. *Oncol. Rep.* **2005**, *13*, 329-333.
16. Teixeira, M.J.; de Almeida, Y.M.; Viana, J.R.; Holanda, Filha J.G.; Rodrigues, T.P.; Prata, J.R.; Jr, Coêlho, I.C.; Rao, V.S.; Pompeu, M.M. In vitro and in vivo Leishmanicidal activity of 2-hydroxy-3-(3-methyl-2-butenyl)-1,4-naphthoquinone (lapachol). *Phytother Res.* **2001**, *15*(1), 44-8. [https://doi.org/10.1002/1099-1573\(200102\)15:1<44::aid-pt685>3.0.co;2-1](https://doi.org/10.1002/1099-1573(200102)15:1<44::aid-pt685>3.0.co;2-1).
17. Lopes, J.N.; Cruz, F.S.; Docampo, R.; Vasconcellos, M.E.; Sampaio, M.C.R.; Pinto, A.V.; Gilbert, B. *In vitro* and *in vivo* evaluation of the toxicity of 1,4-naphthoquinone and 1,2-naphthoquinone derivatives against *Trypanosoma cruzi*. *Ann. Trop. Med. Parasitol.* **1978**, *72*, 523-531, <https://doi.org/10.1080/00034983.1978.11719356>.
18. Silva, T.M.S.; Camara, C.A.; Barbosa, T.P.; Soares, A.Z.; da Cunha, L.C.; Pinto, A.C.; Vargas, M.D. Molluscicidal activity of synthetic lapachol amino and hydrogenated derivatives. *Bioorg. Med. Chem.* **2005**, *13*, 193-196, <https://doi.org/10.1016/j.bmc.2004.09.043>.
19. Oliveira, C.G.T.; Miranda, F.F.; Ferreira, V.F.; Freitas, C.C.; Rabello, R.F.; Carballido, J.M.; Corrêa, L.C.D. Synthesis and antimicrobial evaluation of 3-hydrazino-naphthoquinones as analogs of lapachol. *J. Braz. Chem. Soc.* **2001**, *12*, 339-345, <https://doi.org/10.1590/S0103-50532001000300004>.
20. Jain, M. Assessment of bioactivity of some chemical markers from *feronia limonia* and *tecomella undualta* used in traditional medicines. PhD Thesis, Faculty of Technology and Engineering, Maharaja Sayajirao University of Baroda, Gujarat, India; september **2011**.
21. de Almeida, E.R.; da Silva Filho, A.A.; Dos Santos, E.R.; Correia Lopes, C.A. Antiinflammatory action of lapachol. *J. Ethnopharmacol.* **1990**, *29*, 239-241, [https://doi.org/10.1016/0378-8741\(90\)90061-w](https://doi.org/10.1016/0378-8741(90)90061-w).
22. Borris, R.P.; Schaeffer, J.M. Antiparasitic agents from plants, in *Phytochemical Resources for Medicine and Agriculture* (H. N. Nigg and D. S. Seigler, eds.), Plenum Press, New York **1992**, 117-158.
23. Rodrigues, A.M.S.; de Paula, J.E.; Roblot, F.; Fournet, A.; Espíndola, L.S. Larvicidal activity of *Cydistax antisiphilitica* against *Aedes aegypti* larvae. *Fitoterapia* **2005**, *76*, 755-757, <https://doi.org/10.1016/j.fitote.2005.08.015>.
24. Carvalho, L.H.; Rocha, E.M.; Raslan, D.S.; Oliveira, A.B.; Krettli, A.U. In vitro activity of natural and synthetic naphthoquinones against erythrocytic stages of *Plasmodium falciparum*. *Braz. J. Med. Biol. Res.* **1988**, *21*, 485-487.
25. Portillo, A.; Vila, R.; Freixa, B.; Adzet, T.; Cañigüeral, S. Antifungal activity of Paraguayan plants used in traditional medicine. *J. Ethnopharmacol.* **2001**, *76*, 93-98, [https://doi.org/10.1016/s0378-8741\(01\)00214-8](https://doi.org/10.1016/s0378-8741(01)00214-8).
26. Pereira, E.M.; Machado, T.d.B.; Leal, I.C.R.; Jesus, D.M.; Damaso, C.R.d.A.; Pinto, A.V.; Giambiagi-deMarval, M.; Kuster, R.M.; dos Santos, K.R.N. *Tabebuia avellaneda* naphthoquinones: activity against methicillin-resistant staphylococcal strains, cytotoxic activity and *in vivo* dermal irritability analysis. *Ann. Clin. Microbiol. Antimicrob.* **2006**, *5*, 5, <https://doi.org/10.1186/1476-0711-5-5>.
27. Ventura Pinto, A.; Lisboa de Castro, S. The Trypanocidal Activity of Naphthoquinones: A Review. *Molecules* **2009**, *14*, 4570-4590, <https://doi.org/10.3390/molecules14114570>.
28. Goel, R.K.; Pathak, N.K.R.; Biswas, M.; Pandey, V.B.; Sanyal, A.K. Effect of lapachol, a naphthaquinone isolated from *Tectona grandis*, on experimental peptic ulcer and gastric secretion. *J. Pharm. Pharmacol.* **1987**, *39*, 138-140, <https://doi.org/10.1111/j.2042-7158.1987.tb06962.x>.

29. Li, C.J.; Zhang, L.J.; Dezube, B.J.; Crumpacker, C.S.; Pardee, A.B. Three inhibitors of type 1 human immunodeficiency virus long terminal repeat-directed gene expression and virus replication. *Proc. Natl. Acad. Sci.* **1993**, *90*, 1839-1842, <https://doi.org/10.1073/pnas.90.5.1839>.
30. Dinnen, R.D.; Ebisuzaki, K. The search for novel anticancer agents: a differentiation-based assay and analysis of a folklore product. *Anticancer Res.* **1997**, *17*, 1027-1033.
31. Hussain, H.; Krohn, K.; Ahmad, V.U.; Miana, G.A.; Green, I.R. Lapachol: an overview. *Arkivoc* **2007**, 145-171.
32. Goswami, D.V.; Nirmal, S.A., Patil, M.J.; Dighe, N.S.; Laware, R.B.; Pattan, S.R. An Overview of *Tectona grandis*: Chemistry and Pharmacological Profile. *Phcog Rev.* **2009**, *3*(5), 181-185.
33. Faix, O.; Meier, D.; Grobe, I. Studies on isolated lignins and lignins in woody materials by pyrolysis-GCMS and off-line pyrolysis-GC with flame ionization detection. *Journal of Analytical and Applied Pyrolysis* **1987**, *11*, 403-416.
34. Otten, S.L.; Rosazza, J.P. Oxidative ring fission of the Naphthoquinones lapachol and dichloroallyl lawsone by *Penicillium notatum*. *J. Biol. Chem.* **1983**, *258*, 1610-1613.
35. Khera, N.; Bhargava, S. PHYTOCHEMICAL AND PHARMACOLOGICAL EVALUATION OF *TECTONA GRANDIS*. LINN. *Int. J. Pharm. Pharm. Sci.* **2013**, *5*, 923-927.
36. Khan, R.M.; Mlungwana, S.M. 5-Hydroxylapachol: a cytotoxic agent from *Tectona grandis*. *Phytochemistry* **1999**, *50*, 439-442, [https://doi.org/10.1016/S0031-9422\(98\)00478-6](https://doi.org/10.1016/S0031-9422(98)00478-6).
37. Verma, K.S.; Jain, A.K.; Gupta, S.R. Structure of Undulatin: A New Iridoid Glucoside from *Tecomella undulata*. *Planta Med.* **1986**, *52*, 359-362, <https://doi.org/10.1055/s-2007-969184>.
38. Singh, P.; Khandelwal, P.; Hara, N.; Asai, T.; Fujimoto, Y. Radermachol and naphthoquinone derivatives from *Tecomella undulata*: Complete ¹H and ¹³C NMR assignments of radermachol with the aid of computational ¹³C shift prediction. *Indian J. Chem.* **2008**, *47B*, 1865-1870.
39. FAO. 2020. Global Forest Resources Assessment 2020: Main report. Rome; **2020**; <https://doi.org/10.4060/ca9825en>.
40. Kiranmai, G.; Koundinya, K. ANTIBACTERIAL ACTIVITY OF *TECOMASTANS* AGAINST GRAM POSITIVE AND GRAM NEGATIVE BACTERIA. *IAJPS* **2014**, *1*, 30-34, <https://doi.org/10.5281/zenodo.2648364>.
41. Tomlinson, P.B.; Tomlinson, P.B. The Botany of Mangroves; Cambridge University Press: **1994**.
42. Shetgiri, N.P.; Kokitkar, S.V.; Sawant, S.N. Radermachera xylocarpa: the highly efficient source of lapachol and synthesis of its derivatives. *Acta Pol. Pharm.* **2001**, *58*, 133-135.
43. Cragg, G.M.; Newman, D.J. Plants as a source of anticancer drugs. *J. Ethanopharmacol.* **2005**, *100*, 72-79, <https://doi.org/10.1016/j.jep.2005.05.011>.
44. Kishore, N.; Mishra, B.B.; Tiwari, V.K.; Tripathi, V. Difuranonaphthoquinones from *Plumbago zeylanica* roots. *Phytochem. Lett.* **2010**, *3*, 62-65, <https://doi.org/10.1016/j.phytol.2009.11.007>.
45. Binutu, O.A.; Adesogan, K.E.; Okogun, J.I. Antibacterial and Antifungal Compounds from *Kigelia pinnata*. *Planta Med.* **1996**, *62*, 352-353, <https://doi.org/10.1055/s-2006-957900>.
46. Sadananda, T.S.; Nirupama, R.; Chaithra, K.; Govindappa, M.; Chandrappa, C.P.; Vinay Raghavendra, B. Antimicrobial and antioxidant activities of endophytes from *Tabebuia argentea* and identification of anticancer agent (lapachol). *J. Med. Plants Res.* **2011**, *5*, 3643-3652.
47. Jassbi, A.R.; Singh, P.; Jain, S.; Tahara, S. Novel Naphthoquinones from *Heterophragma adenophyllum*. *Helv. Chim. Acta* **2004**, *87*, 820-824, <https://doi.org/10.1002/hlca.200490080>.
48. Farzana, M.U.Z.N.; Tharique, I.A.; Sultana, A. A review of ethnomedicine, phytochemical and pharmacological activities of *Acacia nilotica* (Linn) willd. *J. Pharmacog. Phytochem.* **2014**, *3*, 84-90.
49. Araújo, I.A.C.; de Paula, R.C.; Alves, C.L.; Faria, K.F.; Oliveira, M.M.; Mendes, G.G.; Dias, E.M.F.A.; Ribeiro, R.R.; Oliveira, A.B.; Silva, S.M.D. Efficacy of lapachol on treatment of cutaneous and visceral leishmaniasis. *Exp Parasitol.* **2019**, *199*, 67-73. <https://doi.org/10.1016/j.exppara.2019.02.013>.
50. Fonseca, S.G.; da C., Braga Carvalho, R.M.; Santana, D.P. Lapachol-Chemistry, pharmacology and assay methods. *Rev Bras Farm* **2003**, *84*(1), 9-16.
51. Lira, A.A.M.; Sester, E.A.; Carvalho, A.L.M.; Strattmann, R.R.; Albuquerque, M.M.; Wanderley, A.G.; Santana, D.P. Development of Lapachol Topical Formulation: Anti-inflammatory Study of a Selected Formulation. *AAPS PharmSciTech* **2008**, *9*, 163-168, <https://doi.org/10.1208/s12249-007-9002-z>.
52. Finkel, J.M.; Harrison, S.D. Fluorometric method for the determination of lapachol in serum. *Anal. Chem.* **1969**, *41*, 1854-1855, <https://doi.org/10.1021/ac60282a035>.

53. Aucélio, R.Q.; Pérez-Cordovés, A.I.; Xavier Lima, J.L.; Ferreira, A.B., Esteva Guas, A.M.; da Silva, A.R. Determination of lapachol in the presence of other naphthoquinones using 3MPA-CBTe quantum dots fluorescent probe. *Spectrochim Acta A Mol Biomol Spectrosc.* 2013 Jan 1;100:155-60. <https://doi.org/10.1016/j.saa.2012.04.020>.
54. Juliano, L.; Aurora, Pérez-Gramatges.; Aucelio, Ricardo.; Silva, A. Improved quantum dots fluorescence quenching using organized medium: A study of the effect of naphthoquinones aiming the analysis of plant extracts. *Microchemical Journal.* **2013**, *110*, 775-782, <https://doi.org/10.1016/j.microc.2013.04.009>.
55. Fonseca, S.G.C.; Da Silva, L.B.L.; Castro, R.F.; De Santana, D.P. Validation of the analytical methodology for evaluation of lapachol in solution by HPLC, *Quim. Nova* **2004**, *27*, 157–159, <https://doi.org/10.1590/S0100-40422004000100026>.
56. Aucélio, R.Q.; Pérez-Cordovés, A.I.; Xavier Lima, J.L.; Ferreira, A.B.B.; Esteva Guas, A.M.; da Silva, A.R. Determination of lapachol in the presence of other naphthoquinones using 3MPA-CdTe quantum dots fluorescent probe. *Spectrochim. Acta A Mol. Biomol. Spectrosc.* **2013**, *100*, 155-160, <https://doi.org/10.1016/j.saa.2012.04.020>.
57. Ali, A.; Kiderlen, A.; Kolodziej, H. Lapachol and isomeric 5- and 8-hydroxy-2-(1'-hydroxyethyl)naphtho[2,3-b] furan-4,9-diones are effective antileishmanial constituents of *Tabebuia avellanadae*. *Planta Medica* **2010**, *76*(12), <https://doi.org/10.1055/s-0030-1264769>.
58. Goulart, M.I.O.F.; Falkowski, P.; Ossowski, T.; Liwo, A. Electrochemical study of oxygen interaction with lapachol and its radical anions. *Bioelectrochemistry* **2003**, *59*, 85-87, [https://doi.org/10.1016/s1567-5394\(03\)00005-7](https://doi.org/10.1016/s1567-5394(03)00005-7).
59. Pinto, C.N.; Dantas, A.P.; De Moura, K.C.G.; Emery, F.S.; Polequevitch, P.F.; Maria do Carmo, F.R.P.; de Castro, S.L.; Pinto, A.V. Chemical Reactivity Studies with Naphthoquinones from *Tabebuia* with Anti-trypanosomal Efficacy. *Arzneimittelforschung* **2000**, *50*, 1120-1128, <https://doi.org/10.1055/s-0031-1300337>.
60. Santos, A.F.; Ferraz, P.A.; de Abreu, F.C.; Chiari, E.; Goulart, M.O.; Sant'Ana, A.E. Molluscicidal and trypanocidal activities of lapachol derivatives. *Planta Med.* **2001**, *67*(1), 92-93, <https://doi.org/10.1055/s-2001-10877>. PMID: 11270734.
61. Park, B.-S.; Lee, H.-K.; Lee, S.-E.; Piao, X.-L.; Takeoka, G.R.; Wong, R.Y.; Ahn, Y.-J.; Kim, J.-H. Antibacterial activity of *Tabebuia impetiginosa* Martius ex DC (Taheebo) against *Helicobacter pylori*. *J. Ethnopharmacol.* **2006**, *105*, 255-262, <https://doi.org/10.1016/j.jep.2005.11.005>.
62. Souza, M.A.; Johann, S.; Lima, L.A.R.d.S.; Campos, F.F.; Mendes, I.C.; Beraldo, H.; Souza-Fagundes, E.M.d.; Cisalpino, P.S.; Rosa, C.A.; Alves, T.M.d.A. The antimicrobial activity of lapachol and its thiosemicarbazone and semicarbazone derivatives. *Mem. Inst. Oswaldo Cruz* **2013**, *108*, 342-351, <https://doi.org/10.1590/S0074-02762013000300013>.
63. Guiraud, P.; Steiman, R.; Campos-Takaki, G.-M.; Seigle-Murandi, F.; de Buochberg, M.S. Comparison of Antibacterial and Antifungal Activities of Lapachol and β -Lapachone. *Planta Med.* **1994**, *60*, 373-374, <https://doi.org/10.1055/s-2006-959504>.
64. Howland, J.L. The reversibility of inhibition by 2-heptyl-4-hydroxyquinolin-*N*-oxide and 2-hydroxy-3-(3-methylbutyl)-1,4-naphthoquinone of succinate oxidation. *Biochim. Biophys. Acta - Gen. Subj.* **1963**, *73*, 665-667, [https://doi.org/10.1016/0006-3002\(63\)90343-3](https://doi.org/10.1016/0006-3002(63)90343-3).
65. Sadananda, T.S.; Nirupama, R.; Chaithra, K.; Govindappa, M.; Chandrappa, C.P.; Raghavendra, V.B. Antimicrobial and antioxidant activities of endophytes from *Tabebuia argentea* and identification of anticancer agent (lapachol). *J. Med. Plants Res.* **2011**, *5*, 3643-3652, <https://doi.org/10.5897/JMPR.9000170>.
66. Schuerch, A.R.; Wehrli, W. β -Lapachone, an Inhibitor of Oncornavirus Reverse Transcriptase and Eukaryotic DNA Polymerase- α . *Eur J of Biochem* **1978**, *84*(1), 197-205, <https://doi.org/10.1111/j.1432-1033.1978.tb12157.x>.
67. Schuerch, A.; Wehrli, W. β -Lapachone, an Inhibitor of Oncornavirus Reverse Transcriptase and Eukaryotic DNA Polymerase- α . *Eur. J. Biochem.* **1978**, *84*, 197-205, <https://doi.org/10.1111/j.1432-1033.1978.tb12157.x>.
68. Junior, N.N.; Santos, I.A.; Meireles, B.A.; Nicolau, M.S.P.; Lapa, I.R.; Aguiar, R.S.; Jardim, A.C.G.; José, D.P. In silico evaluation of lapachol derivatives binding to the Nsp9 of SARS-CoV-2. *J. Biomol. Struct. Dyn.* **2022**, *40*, 5917–5931, <https://doi.org/10.1080/07391102.2021.1875050>.
69. Preusch, P.C.; Suttie, J.W. Lapachol inhibition of vitamin K epoxide reductase and vitamin K quinone reductase. *Arch. Biochem. Biophys.* **1984**, *234*, 405-412, [https://doi.org/10.1016/0003-9861\(84\)90286-8](https://doi.org/10.1016/0003-9861(84)90286-8).

70. Dvorkin-Camiel, L.; Whelan, J.S. Tropical American Plants in the Treatment of Infectious Diseases. *J of Dietary Suppl.* **2008**, *5*(4), 349–372, <https://doi.org/10.1080/19390210802519648>.
71. Ueda, S.; Umemura, T.; Dohguchi, K.; Matsuzaki, T.; Tokuda, H.; Nishino, H.; Iwashima, A. Production of anti-tumour-promoting furano-naphthoquinones in *Tabebuia avellaneda* cell cultures. *Phytochem* **1994**, *36*(2), 323-325, [https://doi.org/10.1016/S0031-9422\(00\)97069-9](https://doi.org/10.1016/S0031-9422(00)97069-9).
72. Wuerzberger, S.M.; Pink, J.J.; Planchon, S.M.; Byers, K.L.; Bornmann, W.G.; Boothman, D.A. Induction of apoptosis in MCF-7:WS8 breast cancer cells by beta-lapachone. *Cancer Res.* **1998**, *58*, 1876-1885.
73. Murray, M.T.; Pizzorno, J.E. *Encyclopedia of Natural Medicine*, 2nd Edition; PA4 Rocklin, Prima Pub: **1998**; pp. 967-972.
74. Kumagai, Y.; Tsurutani, Y.; Shinyashiki, M.; Homma-Takeda, S.; Nakai, Y.; Yoshikawa, T.; Shimojo, N. Bioactivation of lapachol responsible for DNA scission by NADPH-cytochrome P450 reductase. *Environ. Toxicol. Pharmacol.* **1997**, *3*, 245-250, [https://doi.org/10.1016/s1382-6689\(97\)00019-7](https://doi.org/10.1016/s1382-6689(97)00019-7).
75. de Andrade, J.K.F.; da Silva Góes, A.J.; Barbosa, V.X.; de Lima Silva, M.S.; Matos Donato, M.A.; Peixoto, C.A.; Militão, G.C.G.; da Silva, T.G. Anticancer activity of β -Lapachone derivatives on human leukemic cell lines. *Chem.-Biol. Interact.* **2022**, *365*, 110057, <https://doi.org/10.1016/j.cbi.2022.110057>.
76. Jung, E.J.; Kim, H.J.; Shin, S.C.; Kim, G.S.; Jung, J.-M.; Hong, S.C.; Kim, C.W.; Lee, W.S. *Artemisia annua* L. Polyphenols Enhance the Anticancer Effect of β -Lapachone in Oxaliplatin-Resistant HCT116 Colorectal Cancer Cells. *Int. J. Mol. Sci.* **2023**, *24*, 17505, <https://doi.org/10.3390/ijms242417505>.
77. Costa, W.F.; Oliveira, A.B.; Nepomuceno, J.C. Lapachol as an epithelial tumor inhibitor agent in *Drosophila melanogaster* heterozygote for tumor suppressor gene *wts*. *Genet. Mol. Res.* **2011**, *10*, 3236-3245, <https://doi.org/10.4238/2011.December.22.1>.
78. Hamid, F. Molecular Docking and *In-silico* Studies of Lapachol and 2-(1-hydroxyethyl)-naphtho[2,3-b]furan-4,9-dione Against Human Estrogen Receptor Alpha. *Med. Chem.* **2023**, *13*, 1-4.
79. Li, Y.; Feng, M.; Guo, T.; Wang, Z.; Zhao, Y. Tailored Beta-Lapachone Nanomedicines for Cancer-Specific Therapy. *Adv. Healthcare Mater.* **2023**, *12*, 2300349, <https://doi.org/10.1002/adhm.202300349>.
80. Sflakidou, E.; Leonidis, G.; Foroglou, E.; Siokatas, C.; Sarli, V. Recent Advances in Natural Product-Based Hybrids as Anti-Cancer Agents. *Molecules* **2022**, *27*, 6632, <https://doi.org/10.3390/molecules27196632>.
81. De Grandis, R.A.; Oliveira, K.M.; Guedes, A.P.M.; dos Santos, P.W.S.; Aissa, A.F.; Batista, A.A.; Pavan, F.R. A Novel Ruthenium(II) Complex With Lapachol Induces G2/M Phase Arrest Through Aurora-B Kinase Down-Regulation and ROS-Mediated Apoptosis in Human Prostate Adenocarcinoma Cells. *Front. Oncol.* **2021**, *11*, 682968, <https://doi.org/10.3389/fonc.2021.682968>.
82. Atolani, O.; Olatunji, G.A.; Bello, O.M. In vitro anticancer potential of lapachol and derivatized conjugates from *Kigelia pinnata*. *Planta Med.* **2015**, *81*, PB24, <https://doi.org/10.1055/s-0035-1545179>.
83. Ortega, M.J.; Parra-Torrejón, B.; Cano-Cano, F.; Gómez-Jaramillo, L.; González-Montelongo, M.C.; Zubía, E. Synthesis and Antioxidant/Anti-Inflammatory Activity of 3-Arylphthalides. *Pharmaceuticals* **2022**, *15*, 588, <https://doi.org/10.3390/ph15050588>.
84. Basit, A.; Ahmad, S.; Khan, K.u.R.; Aati, H.Y.; Sherif, A.E.; Ovatlarnporn, C.; Khan, S.; Rao, H.; Arshad, M.A.; Shahzad, M.N.; Perveen, S. Evaluation of the anti-inflammatory, antioxidant, and cytotoxic potential of *Cardamine amara* L. (Brassicaceae): A comprehensive biochemical, toxicological, and *in silico* computational study. *Front. Chem.* **2023**, *10*, 1077581, <https://doi.org/10.3389/fchem.2022.1077581>.
85. Nayak, S.S.; Naidu, A.; Sudhakaran, S.L.; Vino, S.; Selvaraj, G. Prospects of Novel and Repurposed Immunomodulatory Drugs against Acute Respiratory Distress Syndrome (ARDS) Associated with COVID-19 Disease. *J. Pers. Med.* **2023**, *13*, 664, <https://doi.org/10.3390/jpm13040664>.
86. Trinh, X.-T.; Long, N.-V.; Van Anh, L.T.; Nga, P.T.; Giang, N.N.; Chien, P.N.; Nam, S.-Y.; Heo, C.-Y. A Comprehensive Review of Natural Compounds for Wound Healing: Targeting Bioactivity Perspective. *Int. J. Mol. Sci.* **2022**, *23*, 9573, <https://doi.org/10.3390/ijms23179573>.
87. Akbari, F.; Azadbakht, M.; Bagheri, A.; Vahedi, L. *In Silico*, *In Vitro*, and *In Vivo* Wound Healing Activity of *Astragalus microcephalus* Willd. *Adv. Pharmacol. Pharm. Sci.* **2022**, *2022*, 2156629, <https://doi.org/10.1155/2022/2156629>.
88. Rauf, A.; AlOmar, T.S.; Sarfaraz, S.; Ayub, K.; Hussain, F.; Rashid, U.; Almasoud, N.; AlOmar, A.S.; Rehman, G.; Ahmad, Z.; Muhammad, N.; Shah, Z.A.; Formanowicz, D. Density functional theory, molecular docking, *In vitro* and *In vivo* anti-inflammatory investigation of lapachol isolated from *Fernandoa adenophylla*. *Heliyon* **2023**, *9*, e22575, <https://doi.org/10.1016/j.heliyon.2023.e22575>.
89. Santos, T. Therapeutic Applications and Adaptability of Multifaceted Medicine Lapachol. *Der Pharmacia Lettre.* **2023**, *15*, 19-20.

90. Bibi, S., Ahmad, F., Alam, M. R., Ansar, M., Yeou, K. S., & Wahedi, H. M. Lapachol-Induced Upregulation of Sirt1/Sirt3 is linked with Improved Skin Wound Healing in Alloxan-induced Diabetic Mice. *Iran. J. Pharm. Sci.* **2021**, *20*, 419. <https://doi.org/10.22037/ijpr.2021.112722.13914>
91. Matsui, T., Ito, C., Oda, M., Itoigawa, M., Yokoo, K., Okada, T., & Furukawa, H. Lapachol suppresses cell proliferation and secretion of interleukin-6 and plasminogen activator inhibitor-1 of fibroblasts derived from hypertrophic scars. *J. Pharm. Pharmacology*, **2011**, *63*, 960-966. <https://doi.org/10.1111/j.2042-7158.2011.01292.x>
92. Sharma, U.K.; Singh, A.; Sharma, U.; Kumar, M.; Rai, D.; Agrahari, P. Wound healing activity of kigelia pinnata bark extract. *Asian J. Pharm. Clin. Res.* **2010**, *3*, 73-75.
93. Shi, X.; Yang, J.; Deng, S.; Xu, H.; Wu, D.; Zeng, Q.; Wang, S.; Hu, T.; Wu, F.; Zhou, H. TGF- β signaling in the tumor metabolic microenvironment and targeted therapies. *J. Hematol. Oncol.* **2022**, *15*, 135, <https://doi.org/10.1186/s13045-022-01349-6>.
94. Freitas, C.S.; Pereira, P.R.; Cardoso, R.V.; Pauli, F.P.; Ribeiro, R.C.B.; Da Silva, F.D.C.; Ferreira, V.F.; Paschoalin, V.M.F. Antimicrobial and wound healing potential of naphthoquinones encapsulated in nanochitosan. *Front. Bioeng. Biotechnol.* **2024**, *11*, 1284630, <https://doi.org/10.3389/fbioe.2023.1284630>.
95. Morrison, R.K.; Brown, D.E.; Oleson, J.J.; Cooney, D.A. Oral toxicology studies with Lapachol. *Toxicol. Appl. Pharmacol.* **1970**, *17*, 1-11, [https://doi.org/10.1016/0041-008x\(70\)90126-2](https://doi.org/10.1016/0041-008x(70)90126-2).
96. Felício, A.C.; Chang, C.V.; Brandão, M.A.; Peters, V.M.; Guerra, M.d.O. Fetal growth in rats treated with lapachol. *Contraception* **2002**, *66*, 289-293, [https://doi.org/10.1016/s0010-7824\(02\)00356-6](https://doi.org/10.1016/s0010-7824(02)00356-6).
97. Strauch, M.A.; Tomaz, M.A.; Monteiro-Machado, M.; Cons, B.L.; Patrão-Neto, F.C.; Teixeira-Cruz, J.D.M.; Tavares-Henriques, M.D.S.; Nogueira-Souza, P.D.; Gomes, S.L.S.; Costa, P.R.R.; Schaeffer, E.; da Silva, A.J.M.; Melo, P.A. Lapachol and synthetic derivatives: *in vitro* and *in vivo* activities against Bothrops snake venoms. *PLoS One* **2019**, *14*(1), e0211-229.